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### Total Synthesis of Glycohexa- and Nonaosyl Ceramide with a Sialyl Le<sup>a</sup> and Sialyl Dimeric Le<sup>a</sup> Sequence

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**TOTAL SYNTHESIS OF GLYCOHEXA- AND NONAOSYL CERAMIDE  
WITH A SIALYL Le<sup>a</sup> AND SIALYL DIMERIC Le<sup>a</sup> SEQUENCE<sup>1</sup>**

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**ABSTRACT**

A total synthesis of tumor-associated glycolipid antigen sialyl Le<sup>a</sup>, and a first total synthesis of glycononaosyl ceramide with a dimeric Le<sup>a</sup> sequence are described. Regio- and stereoselective coupling of sialyl donor **12** with suitably protected Le<sup>a</sup> trisaccharide fluoride **17** was performed by orthogonal glycosylation to give the expected tetrasaccharide **18** in good yield. Glycosylation of its acetate **19** with lactose derivative **8** by use of Cp<sub>2</sub>HfCl<sub>2</sub>-AgOTf as a promoter, gave the desired hexasaccharide **20**. On the other hand, glycosylation between sialyl Le<sup>a</sup> tetrasaccharide donor **34** and Le<sup>a</sup> pentasaccharide acceptor **35** under the agency of PhSeOTf afforded the desired nonasaccharide **36**. After replacement of the benzyl groups in **20** and **36** by acetyl groups, the anomeric acetate was transformed into the  $\alpha$ -trichloroacetimidates **23**, **39** and the fluoride derivative **40**. The crucial couplings of **23**, **39** and **40** with azidosphingosine derivative **3** or ceramide derivative **4** were executed to afford  $\beta$ -glycosides **24** and **41**. Compound **24** was converted to **26** *via* reduction of the azide group followed by *N*-acylation. The target gangliosides **1** and **2** were obtained from **26**

and **41**, respectively, by selective cleavage of the methyl ester, conversion of the *N*-phthalimido group to the *N*-acetamido group, and *O*-deprotection.

## INTRODUCTION

Sialyl Le<sup>a</sup> ganglioside has been isolated<sup>2</sup> from human adenocarcinoma cell line SW1116, and found<sup>3</sup> to be widespread as the tumor-associated glycolipid antigen of digestive organs. Recently it was reported that MUC1 and CD43 mucins, which have a sialyl Le<sup>a</sup> epitope, can inhibit leukocyte adhesion to E-selectin.<sup>4</sup> In 1993, Kitagawa reported the isolation and characterization of hybrid-type sialyl Le<sup>a</sup>-Le<sup>x</sup> glycosphingolipid, but sialyl dimeric Le<sup>a</sup> glycosphingolipid has not been isolated from nature so far.

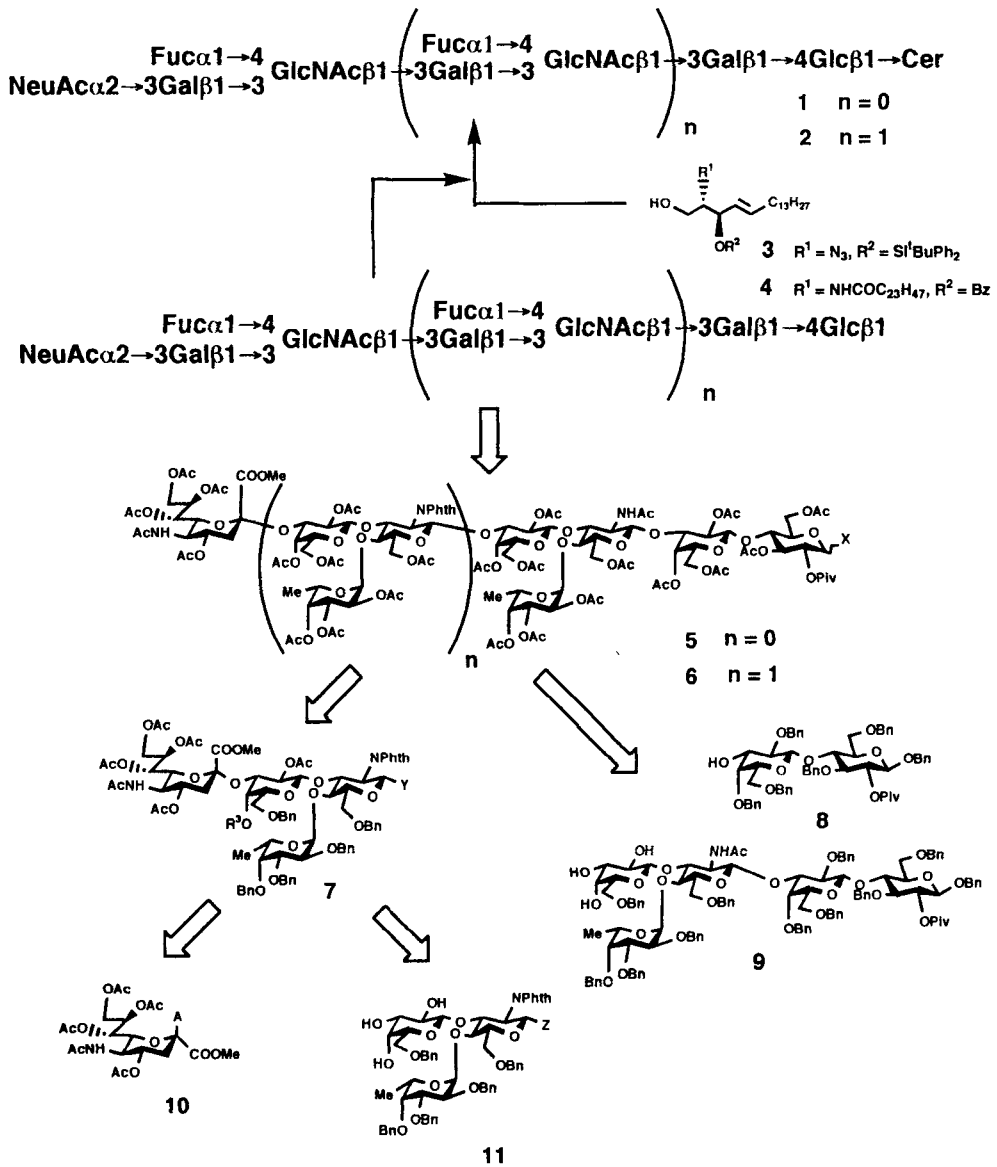
In order to facilitate further biochemical studies of **1** and **2**, an achievement of their efficient synthesis is a challenging task. As part of our current investigation<sup>5-7</sup> on the synthesis of glycosphingolipid with biological importance, we report herein a stereo-controlled synthesis of sialyl Le<sup>a</sup> ganglioside **1** which is carried out by taking a different and efficient strategy from that reported by Hasegawa<sup>8</sup> as well as a first total synthesis of sialyl dimeric Le<sup>a</sup> glycononaosyl ceramide **2**.

## RESULTS AND DISCUSSION

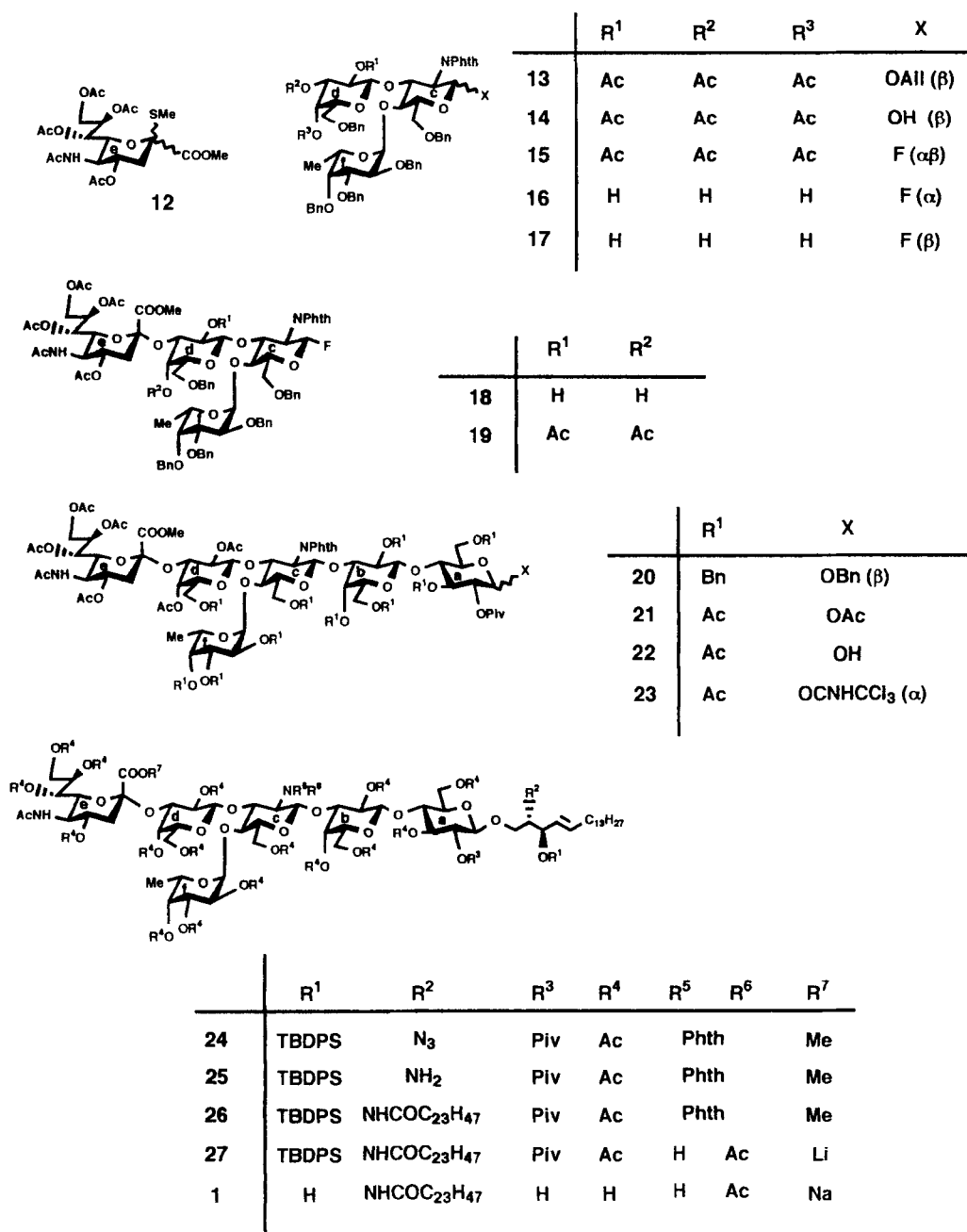
A retrosynthetic analysis of the targets **1**, **2** (Scheme 1) led us to design the putative glycosyl donors **5**, **6** that could be coupled with azidosphingosine derivative **3**<sup>9</sup> or ceramide derivative **4**.<sup>10</sup> The glycosyl donors **5**, **6** were expected to be constructed from the coupling between tetrasaccharide donor **7** and lactose acceptor **8**<sup>11</sup> or pentasaccharide acceptor **9**.<sup>5</sup> The efficiency of the pivaloyl auxiliary group at *O*-2a of **9** was established in previous studies.<sup>12</sup> Putative donor **7** was further dissected into sialic acid donor **10** and Le<sup>a</sup> trisaccharide acceptor **11**.

### 1 Synthesis of SLe<sup>a</sup>-Lac-Cer

In order to construct donor **18**, triol **17**, which was designed to afford a better coupling yield for the next  $\alpha$ -sialylation, was selected as the glycosyl acceptor. Le<sup>a</sup> trisaccharide **13**<sup>5</sup> was converted into compound **17** as follows. Deallylation of **13** with 1) [Ir(COD)(PMeph<sub>2</sub>)<sub>2</sub>]PF<sub>6</sub><sup>13</sup> in THF and 2) I<sub>2</sub> in aq. THF afforded hemiacetal **14** in 97%



Scheme 1



Scheme 2

yield. Hemiacetal **14** was treated with DAST (diethylaminosulfur trifluoride)<sup>14</sup> at -15 °C to give compound **15** in 90% yield ( $\alpha : \beta = 1 : 4$ ). Zemplén *O*-deacetylation of **15** and silica gel separation afforded triol **16** (16%), **17** (71%). To our delight, glycosylation between trisaccharide fluoride **17** and thioglycoside **12**<sup>15</sup> smoothly proceeded under the agency of PhSeOTf<sup>16</sup> in CH<sub>3</sub>CN at -40 °C to give the desired tetrasaccharide **18** and corresponding  $\beta$  epimer in 46% and 14%, respectively, without affecting the anomeric fluoride according to the concept of orthogonal glycosylation strategy.<sup>17</sup> No self-condensed product was detected.

The regiochemistry of newly introduced glycosidic linkage of **18** was deduced by converting **18** into its acetate **19**, which showed in the Homonuclear Hartmann-Hahn (HOHAHA) NMR spectra newly deshielded signals for H-2d at  $\delta$  4.539 and H-4d at  $\delta$  5.142, respectively, confirming that sialyl residue was unambiguously introduced at C-3d of the galactose residue.

The configuration at C-2e in **19** was assigned as  $\alpha$ , based on the <sup>1</sup>H NMR data. The signal for H-4e in **19** was observed at  $\delta$  4.85, and the  $J_{7e,8e}$  value was observed to be 8.4 Hz, consistent with previous observations.<sup>18</sup> Having the designed tetrasaccharide donor **19** in hand, crucial glycosylation was examined. The glycosylation of **19** with lactose derivative **8**<sup>11</sup> was performed in CH<sub>3</sub>CN for 4.5 h at -15 °C in the presence of CP<sub>2</sub>HfCl<sub>2</sub> - AgOTf<sup>19</sup> and molecular sieves 3A, to give the desired hexasaccharide **20** in 65% yield. The configuration of C-1c was expected to be  $\beta$ , due to the presence of the *N*-2 phthaloyl group in **19**, which favors the formation of 1,2-*trans* stereochemistry. Indeed, the <sup>1</sup>H NMR spectrum showed a signal for the anomeric proton of H-1c at  $\delta$  5.152, thus confirming the  $\beta$  configuration.

The transformation of **20** into glycosyl donor **23** was performed as follows. Catalytic hydrogenolysis of **20** by Perlman's catalyst in MeOH-H<sub>2</sub>O (4 : 1) for 17 h, and subsequent acetylation afforded **21** in 99% yield. Chemoselective cleavage of the anomeric acetate of **21** with hydrazinium acetate<sup>20</sup> in DMF afforded hemiacetal **22** in 94% yield. Treatment of **22** with trichloroacetonitrile<sup>21</sup> in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) gave  $\alpha$ -trichloroacetimidate **23** in 91% yield. Significant signals in the <sup>1</sup>H NMR spectrum of **23** were a one-proton doublet at  $\delta$  5.217 ( $J$  3.3Hz) and a one-proton singlet at  $\delta$  8.613 (C = NH), indicating the  $\alpha$ -configuration. The final glycosylation between **23** and **3** was achieved in CH<sub>2</sub>Cl<sub>2</sub> in the presence of boron trifluoride etherate<sup>22</sup> at -15 °C to afford the desired **24** in 86% yield.

The newly introduced glycosidic linkage of **24** was rigorously confirmed to be  $\beta$  ( $\delta_{\text{H}} 4.172$ , d,  $J = 8.1$  Hz), as revealed in the HOHAHA experiment.

Selective reduction of the azide group in **24** with  $\text{Ph}_3\text{P}$  in toluene- $\text{H}_2\text{O} = 2 : 1$ ,<sup>23</sup> and subsequent condensation with tetracosanoic acid, using 2-chloro-1-methylpyridinium iodide and  $\text{Bu}_3\text{N}$  in  $\text{CH}_2\text{Cl}_2$ ,<sup>24</sup> gave the acylated sialyl  $\text{Le}^{\text{a}}$  ganglioside **26** in 88% yield.

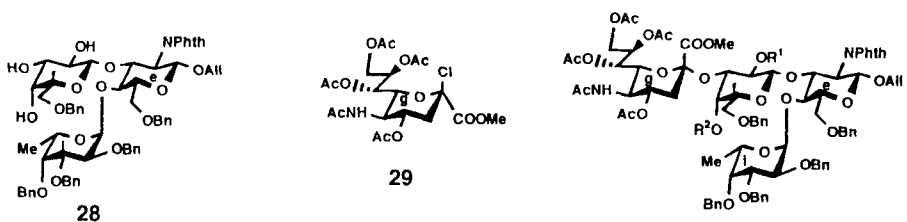
Further conversion into the target glycolipid **1** was executed as follows. Compound **26** was refluxed for 1 day with a large excess of LiI in pyridine<sup>25</sup> to give a 79% yield of the lithium salt **27**. Subsequent treatment of **27** with i)  $\text{NH}_2\text{NHMe}$  in refluxing  $\text{EtOH}$ <sup>26</sup> ii)  $\text{Ac}_2\text{O}$  in  $\text{MeOH}$  iii) aq.  $\text{NaOH}$  in 1 : 1  $\text{MeOH-THF}$  iv)  $\text{Bu}_4\text{NF}$  in  $\text{THF}$ , afforded the target compound **1** in 59% yield, after gel filtration through Sephadex LH-20 using 5 : 5 : 1  $\text{CHCl}_3\text{-MeOH-H}_2\text{O}$ . The  $^1\text{H}$  NMR data of **1** was consistent with the structure assigned. (Figure 1)

## 2 Synthesis of $\text{SdLe}^{\text{a}}\text{-Lac-Cer}$

Glycosylation between three molar equivalent **29**<sup>27</sup> with  $\text{Le}^{\text{a}}$  trisaccharide acceptor **28** in  $\text{CH}_3\text{CN}$  was performed by the action of  $\text{HgBr}_2\text{-Hg(CN)}_2$  at room temperature<sup>28</sup> to afford the desired  $\alpha$ -(2  $\rightarrow$  3)-linked tetrasaccharide **30** in 71% yield based on acceptor, accompanied by a 20% yield of its  $\beta$ -lactone compound. The structure of **31** was confirmed as follows. Acetylation of **30** with acetic anhydride in pyridine at room temperature for 5 h gave only 2-*O*-acetylated **31** in 94% yield. It is worthy of mention that H-4f was not acetylated, presumably due to steric hindrance. HOHAHA NMR spectra showed that H-2f in **31** was deshielded to  $\delta 4.861$ . The configuration at C-2g in compound **31** was assigned as  $\alpha$ ,<sup>29</sup> based on the  $^1\text{H}$  NMR data. The signal for H-4g in **31** was observed at  $\delta 4.74$ , and the  $J_{7g,8g}$  value was observed to be 8.8 Hz,<sup>18</sup> consistent with previous observations.

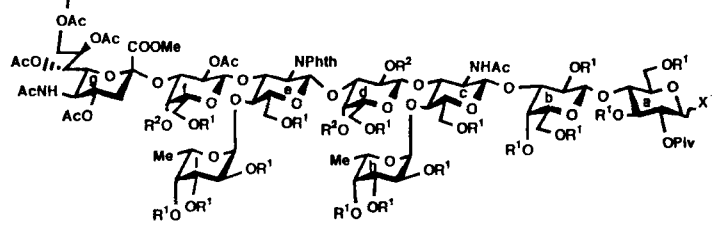
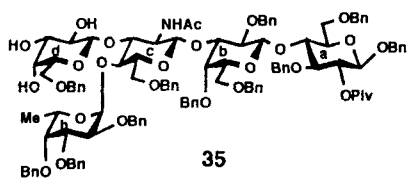
Compound **31** was converted into thioglycoside **34** as follows. Deallylation of **31** was performed as described for **13** to give hemiacetal **32** in quantitative yield. **32** was transformed into  $\beta$ -trichloroacetimidate **33** in 89% yield in the presence of  $\text{CCl}_3\text{CN}$  and 1,8-diaza-bicyclo[5.4.0]undec-7-one (DBU).<sup>21</sup> The imidate **33** was treated with  $\text{Bu}_3\text{SnSMe}$  and boron trifluoride etherate at  $-10^\circ\text{C}$  to give thioglycoside **34** in 96% yield.

Having prepared the designed tetrasaccharide donor **34**, and the pentasaccharide acceptor **35**, crucial glycosylation was examined.  $\text{PhSeOTf}$ -promoted<sup>16</sup> glycosylation between **34** and **35** in  $\text{CH}_3\text{CN}$  at  $-10^\circ\text{C}$  was performed in a regio- and

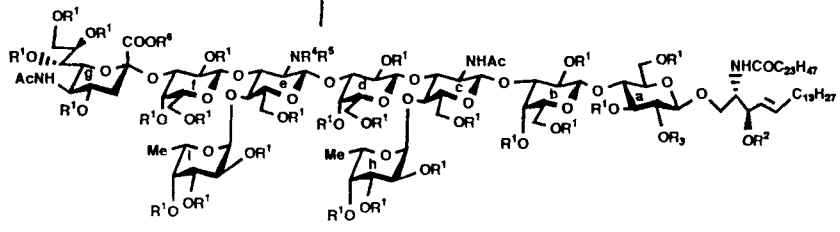


	R <sup>1</sup>	R <sup>2</sup>
30	H	H
31	Ac	H

	X
32	OH
33	OCNHCCl <sub>3</sub>
34	SMe



	R <sup>1</sup>	R <sup>2</sup>	X
36	Bn	H	OBn (β)
37	Ac	Ac	OAc
38	Ac	Ac	OH
39	Ac	Ac	OCNHCCl <sub>3</sub> (α)
40	Ac	Ac	F



	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	R <sup>5</sup>	R <sup>6</sup>
41	Ac	Bz	Piv	Phth	Me	
42	Ac	Bz	Piv	Phth	Li	
2	H	H	H	H	Ac	Na

Scheme 3

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stereo-controlled manner to afford nonasaccharide **36** in 47% yield. (Glycosylation of **33** with **35** in the presence of boron trifluoride etherate at  $-15\text{ }^{\circ}\text{C}$  afforded a 15% yield of **36**). In the  $^1\text{H}$  NMR spectrum of **36**, the anomeric proton of H-1e appeared as a one-proton doublet ( $J = 7.7\text{ Hz}$ ) at  $\delta$  5.234, showing the newly formed glycosidic linkage to be  $\beta$ .

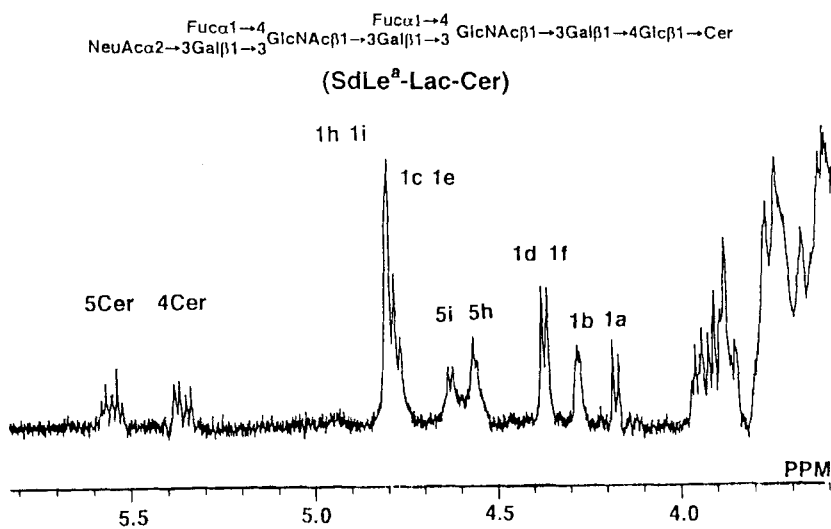
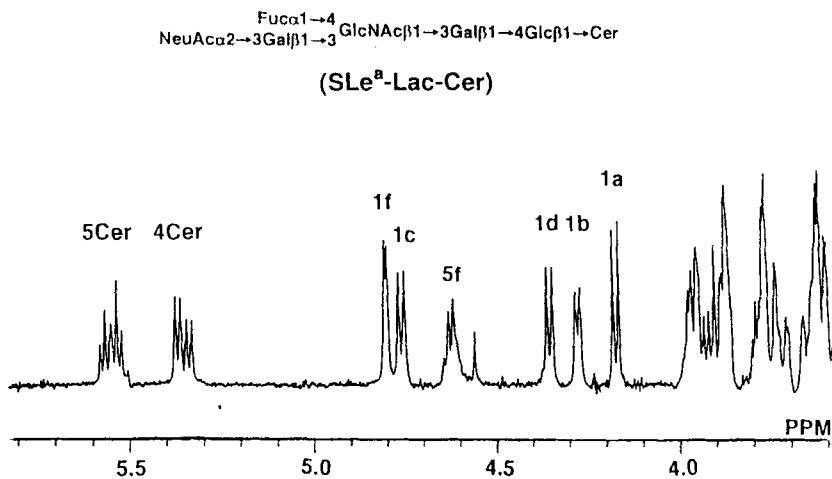
The transformation of **36** into glycosyl donors **39** and **40** was performed as follows. Hydrogenolysis of **36**, subsequent acetylation, and chemoselective cleavage of the anomeric acetate,<sup>20</sup> afforded  $\alpha$ -trichloroacetimidate **39** in 62% yield. The final coupling between **39** and ceramide derivative **4**<sup>9</sup> was achieved in freshly distilled  $\text{CHCl}_3$  in the presence of boron trifluoride etherate<sup>22</sup> at  $-15\text{ }^{\circ}\text{C}$  to afford a 34% yield of the desired **41** accompanied by recovered hemiacetal **38**. The HOHAHA NMR spectral data showed the signals for H-2f and H-4f at  $\delta$  4.804,  $\delta$  4.948 and, H-2d and H-4d at  $\delta$  4.966,  $\delta$  5.246 respectively, indicating the internal dimeric  $\text{Le}^a$  linkages. The newly introduced glycosidic linkage of **41** was rigorously confirmed to be  $\beta$  ( $\delta\text{H}$ , 4.406, d,  $J = 8.1\text{ Hz}$ ). Next aiming at increasing the coupling yield, compound **38** was converted into fluoride **40**. Glycosylation of **40** with ceramide derivative **4** in the presence of  $\text{SnCl}_2\text{-AgOTf}$ <sup>30</sup> at  $-15\text{ }^{\circ}\text{C}$  afforded a 40% yield of **41**.

Further conversion into the target glycolipid **2** was executed as follows. Compound **41** was refluxed for 1 day with a large excess LiI in pyridine<sup>25</sup> to give an 88% yield of the lithium salt **42**. Subsequent treatment of **42** with i)  $\text{NH}_2\text{NHMe}$  in refluxing  $\text{EtOH}$ <sup>26</sup> ii)  $\text{Ac}_2\text{O}$  in 2 : 2 : 1  $\text{MeOH-THF-CH}_2\text{Cl}_2$  iii) aq.  $\text{NaOH}$  in 1 : 1  $\text{MeOH-THF}$ , afforded the target compound **2** in 74% yield, after gel filtration through Sephadex LH-20 using 5 : 5 : 1  $\text{CHCl}_3\text{-MeOH-H}_2\text{O}$ . The structure of **2** was confirmed by  $^1\text{H}$  NMR as shown in Figure 1.

In conclusion, the total synthesis of sialyl monomeric  $\text{Le}^a$  and the first total synthesis of sialyl dimeric  $\text{Le}^a$  ganglioside were achieved by employing the trichloroacetimidate donors **33** and **39** and thioglycoside **34** as key glycosyl donors, and the suitably protected lactose and  $\text{Le}^a$  derivatives **8** and **35** as glycosyl acceptors.

## EXPERIMENTAL

**General methods.** Optical rotations were determined for solutions in  $\text{CHCl}_3$  at  $22 \pm 3\text{ }^{\circ}\text{C}$  with a JASCO Model DIP-370 polarimeter, unless otherwise stated. All



**Figure 1.** Expansions of 500MHz <sup>1</sup>H NMR spectrum of ganglioside 1, 2.

reactions were monitored by high-performance thin-layer chromatography on Kieselgel 60 F<sub>254</sub> (Merck) with detection by UV light and/or by charring with 5% sulfuric acid in ethanol. Flash chromatography was performed on columns of Wakogel C-300 (200~300 mesh). <sup>1</sup>H NMR spectra were recorded with a JNM-GX 500 Fourier-transform instrument. The values of δH are expressed in ppm. downfield from internal Me<sub>4</sub>Si, for solutions in CDCl<sub>3</sub> at 25 °C unless otherwise noted. Fast atom bombardment (FAB) and electrospray ionization (ESI) mass spectroscopy were recorded on a Finnigan MAT TSQ 700 triple stage quadrupole mass spectrometer equipped with an Ion Tech FAB gun or electrospray ion source. Powdered molecular sieves (3A or 4A; GL Sciences Inc. Japan) and lithium iodide were heated to 250 °C under vacuum overnight. All reactions except hydrogenation were performed under atmospheres of dry nitrogen. (ClCH<sub>2</sub>)<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, CH<sub>3</sub>CN, EtCN, CHCl<sub>3</sub>, were distilled from CaH<sub>2</sub>.

***O* - (2,3,4-Tri-*O*-acetyl-6-*O*-benzyl-β-D-galactopyranosyl) - (1→3)-*O*-[(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl) - (1→4)] -6-*O*-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranose (14).** To a solution of (1,5-cyclooctadiene) bis(methyldiphenylphosphine) iridium hexafluorophosphate (57 mg, 48 μmol) which was activated by H<sub>2</sub> in THF (25 mL) was added a solution of **13** (595 mg, 0.48 mmol) in THF (10 mL). The mixture was stirred for 2 h at room temperature, then I<sub>2</sub> (730 mg, 2.88 mmol) and H<sub>2</sub>O (13 mL) was added. This was stirred for 1 h at room temperature. The mixture was diluted with CHCl<sub>3</sub>, washed successively with aq. sodium thiosulfate, aq. NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was chromatographed on SiO<sub>2</sub> with 5 : 1 toluene-EtOAc to afford **14** (232 mg, 97%): [α]<sub>D</sub> -19.8° (c 1.0); R<sub>f</sub> 0.14 (3 : 1 toluene-EtOAc); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.346 (d, 1H, J = 3.3 Hz, h-4d), 5.087 (d, 1H, J = 4.0 Hz, H-1f), 4.970 (dd, 1H, J = 8.1, 9.9 Hz, H-2d), 4.837 (dd, 1H, J = 9.2, 10.6 Hz, H-2c), 4.436 (dd, 1H, J = 3.6, 9.5 Hz, H-3d), 4.201 (d, 1H, J = 8.5 Hz, H-1d), 3.986 (t, 1H, J = 9.5 Hz, H-3c), 1.962, 1.841, 1.661 (3s, 9H, 3Ac), 1.274 (d, 3H, J = 6.6 Hz, H-6f).

Anal. Calcd for C<sub>67</sub>H<sub>71</sub>N<sub>1</sub>O<sub>19</sub>: C, 67.38; H, 5.99; N, 1.17. Found: C, 67.10; H, 6.02; N, 1.23.

***O* - (2,3,4-Tri-*O*-acetyl-6-*O*-benzyl-β-D-galactopyranosyl) - (1→3)-*O*-[(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl) - (1→4)] -6-*O*-benzyl-2-deoxy-2-phthalimido-D-glucopyranosyl Fluoride (15).** To a solution of **14** (120 mg, 0.10 mmol) in (ClCH<sub>2</sub>)<sub>2</sub> (1 mL) was added diethylaminosulfur trifluoride (54 μL, 0.40

mmol) at  $-15^{\circ}\text{C}$ . The mixture was diluted with EtOAc, and washed successively with aq.  $\text{NaHCO}_3$ , brine, dried ( $\text{MgSO}_4$ ) and concentrated in vacuo. Chromatography of the residue on  $\text{SiO}_2$  with 3 : 1 toluene-EtOAc afforded **15** (108 mg, 90%) as a 4 : 1 mixture of  $\beta$  and  $\alpha$  anomers:  $R_f$  0.55 (2 : 1 toluene-EtOAc);  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.687 (dd, 0.8H,  $J = 7.5, 54$  Hz, H-1c( $\beta$ )), 5.341 (d, 0.8H,  $J = 3.0$  Hz, H-4d( $\beta$ )), 5.132 (d, 0.8H,  $J = 3.5$  Hz, H-1f( $\beta$ )), 4.171 (d, 0.8H,  $J = 8.5$  Hz, H-1d( $\beta$ )), 3.894 (dd, 1.6H,  $J = 2.5, 10.0$  Hz, H-2f( $\beta$ ) and H-3f( $\beta$ )), 1.981, 1.840, 1.667 (3s, 9H, 3Ac), 1.275 (d, 2.4H,  $J = 6.5$  Hz, H-6f( $\beta$ )).

Anal. Calcd for  $\text{C}_{67}\text{H}_{70}\text{F}_1\text{N}_1\text{O}_{18}$ : C, 67.27; H, 5.90; N, 1.17. Found: C, 67.07; H, 5.94; N, 1.22.

***O* - (6-*O* - Benzyl -  $\beta$  - D - galactopyranosyl) - (1 $\rightarrow$ 3) - *O* - [(2,3,4 - tri-*O* - benzyl -  $\alpha$  - L - fucopyranosyl) - (1 $\rightarrow$ 4)] - 6-*O* - benzyl - 2-deoxy - 2-phthalimido -  $\alpha$  - D - glucopyranosyl Fluoride (**16**), and *O* - (6-*O* - Benzyl -  $\beta$  - D - galactopyranosyl) - (1 $\rightarrow$ 3) - *O* - [(2,3,4 - tri-*O* - benzyl -  $\alpha$  - L - fucopyranosyl) - (1 $\rightarrow$ 4)] - 6-*O* - benzyl - 2-deoxy - 2-phthalimido -  $\beta$  - D - glucopyranosyl Fluoride (**17**). To a solution of **15** (97 mg, 81  $\mu\text{mol}$ ) in 7 : 2 MeOH-( $\text{ClCH}_2$ ) $_2$  (9 mL) was added 0.1N MeONa (400  $\mu\text{L}$ ), and the mixture was stirred for 2 h at room temperature, neutralized with Amberlyst 15E ( $\text{H}^+$ ) resin, and filtered. The filtrate was concentrated in vacuo. Chromatography of the residue on  $\text{SiO}_2$  with 1 : 1 toluene-EtOAc afforded **16** (14 mg, 16%), **17** (53 mg, 61%).**

Compound **16** had:  $[\alpha]_D +3.98^{\circ}$  ( $c$  1.0);  $R_f$  0.21 (1 : 2 toluene-EtOAc);  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.881 (dd, 1H,  $J = 2.2, 53.5$  Hz, H-1c), 5.110 (d, 1H,  $J = 3.3$  Hz, H-1f), 4.533 (d, 1H,  $J = 7.3$  Hz, H-1d), 3.869 (s, 3H, OMe), 1.173 (d, 3H,  $J = 6.2$  Hz, H-6f).

Anal. Calcd for  $\text{C}_{61}\text{H}_{64}\text{F}_1\text{N}_1\text{O}_{15} \cdot 3\text{H}_2\text{O}$ : C, 65.17; H, 6.28; N, 1.25. Found: C, 65.19; H, 6.24; N, 1.22.

Compound **17** had:  $[\alpha]_D -20.5^{\circ}$  ( $c$  1.0);  $R_f$  0.22 (1 : 2 toluene-EtOAc);  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.776 (dd, 1H,  $J = 7.7, 53.9$  Hz, H-1c), 5.112 (d, 1H,  $J = 3.7$  Hz, H-1f), 4.833 (dd, 1H,  $J = 9.2, 11.0$  Hz, H-2c), 4.018 (d, 1H,  $J = 7.7$  Hz, H-1d), 1.097 (d, 3H,  $J = 6.6$  Hz, H-6f).

***O* - (Methyl 5-Acetamido - 4,7,8,9 - tetra-*O* - acetyl - 3,5 - dideoxy - D - glycerol -  $\alpha$  - D - galacto - 2 - nonulopyranosylate) - (2 $\rightarrow$ 3) - *O* - (6-*O* - benzyl -  $\beta$  - D - galactopyranosyl) - (1 $\rightarrow$ 3) - *O* - [(2,3,4 - tri-*O* - benzyl -  $\alpha$  - L - fucopyranosyl) - (1 $\rightarrow$**

**4)]-6-O-benzyl-2-deoxy-2-phthalimido-β-D-glucofuranosyl Fluoride (18).**

A mixture of **12** (64 mg, 123 μmol), **17** (49 mg, 46 μmol) and 3A molecular sieves (300 mg) in 2 : 1 MeCN-EtCN (1.5 mL) was stirred for 9 h at 20 °C under argon. After cooling to -40 °C, a solution of AgOTf (42 mg, 179 μmol) and PhSeCl (31 mg, 179 μmol) in MeCN (1 mL) were added. After stirring for 3 h at -40 °C, the reaction mixture was quenched with Et<sub>3</sub>N, diluted with EtOAc and filtered through a Celite bed. The filtrate was washed with aq. NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was chromatographed firstly on Bio-Beads SX-4 in toluene and then purified on SiO<sub>2</sub> with 2 : 1 EtOAc-toluene afforded **18** (33 mg, 46%) and corresponding β epimer (10 mg, 14%).

Compound **18** had: R<sub>f</sub> 0.26 (3 : 1 CHCl<sub>3</sub>-acetone); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.812 (m, 4H, Phth), 5.786 (dd, 1H, J = 7.7, 53.9 Hz, H-1c), 5.236 (dd, 1H, J = 1.8, 8.1 Hz, H-7e), 5.099 (d, 1H, J = 4.0 Hz, H-1f), 5.056 (m, 1H, H-4e), 4.842 (t, 1H, J = 9.9 Hz, H-2c), 4.086 (d, 1H, J = 9.2 Hz, H-1d), 3.733 (s, 3H, OMe), 2.442 (d, 1H, J = 4.8, 12.8 Hz, H-3eeq), 2.052, 2.047, 2.017, 1.997, 1.878 (5s, 15H, 5Ac), 1.120 (d, 3H, J = 6.6 Hz, H-6f).

Anal. Calcd for C<sub>81</sub>H<sub>91</sub>F<sub>1</sub>N<sub>2</sub>O<sub>27</sub>: C, 63.03; H, 5.94; N, 1.81. Found: C, 63.13; H, 5.91; N, 1.77.

Compound β epimer had: [α]<sub>D</sub> -23.6° (c 1.0); R<sub>f</sub> 0.28 (3 : 1 CHCl<sub>3</sub>-acetone); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.811 (m, 4H, Phth), 5.794 (dd, 1H, J = 7.7, 54.0 Hz, H-1c), 5.142 (m, 1H, H-4e), 5.096 (d, 1H, J = 4.0 Hz, H-1f), 4.081 (d, 1H, J = 9.2 Hz, H-1d), 2.405 (d, 1H, J = 4.6, 12.8 Hz, H-3e eq), 1.122 (d, 3H, J = 6.6 Hz, H-6f).

**O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate) - (2→3)-O-(2,4-di-O-acetyl-6-O-benzyl-β-D-galactopyranosyl) - (1→3)-O-[(2,3,4-tri-O-benzyl-α-L-fucopyranosyl) - (1→4)] - 6-O-benzyl-2-deoxy-2-phthalimido-β-D-glucofuranosyl Fluoride (19).** To a solution of **18** (173 mg, 112 μmol) in pyridine (5 mL) was added acetic anhydride (5 mL), and the mixture was stirred for 5 days at room temperature. Catalytic amount of DMAP was added, and the reaction mixture was stirred for another 5 days at room temperature, and coevaporated with toluene. Chromatography of the residue on SiO<sub>2</sub> with 8 : 1 toluene-MeOH afforded **19** (152 mg, 83%): R<sub>f</sub> 0.50 (5 : 1 toluene-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.807 (m, 4H, Phth), 5.590 (dd, 1H, J = 7.7, 54.6 Hz, H-1c), 5.278 (dd, 1H, J = 2.6, 8.4 Hz, H-7e), 5.142 (d, 1H, J = 3.7 Hz, H-4d or

H-1f), 4.539 (t, 1H,  $J = 8.1$  Hz, H-2d), 3.774 (s, 3H, OMe), 2.452 (dd, 1H,  $J = 4.8, 12.5$  Hz, H-3 $_{eeq}$ ), 2.133, 2.060, 2.049, 1.979, 1.915, 1.818, 1.699 (7s, 21H, 7Ac), 1.321 (d, 3H,  $J = 6.6$  Hz, H-6f).

Anal. Calcd for  $C_{85}H_{95}F_1N_2O_{29}$ : C, 62.72; H, 5.88; N, 1.72. Found: C, 62.55; H, 5.92; N, 1.75.

**Benzyl O - (Methyl - 5 - acetamido - 4, 7, 8, 9 - tetra - O - acetyl - 3, 5 - dideoxy - D - glycerol -  $\alpha$  - D - galacto - 2 - nonulopyranosylate) - (2  $\rightarrow$  3) - O - (2, 4 - di - O - acetyl - 6 - O - benzyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  3) - O - [(2, 3, 4 - tri - O - benzyl -  $\alpha$  - L - fucopyranosyl) - (1  $\rightarrow$  4)] - O - (6 - O - benzyl - 2 - deoxy - 2 - phthalimido -  $\beta$  - D - glucopyranosyl) - (1  $\rightarrow$  3) - (2, 4, 6 - tri - O - benzyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  4) - 3, 6 - di - O - benzyl - 2 - O - pivaloyl -  $\beta$  - D - glucopyranoside (20).** To a stirred mixture of  $Cp_2HfCl_2$  (57 mg, 145  $\mu$ mol), AgOTf (77 mg, 290  $\mu$ mol) and 3A molecular sieves (1.4 g) was added a solution of **19** (135 mg, 83  $\mu$ mol) and **8** (120 mg, 124  $\mu$ mol) in  $CH_3CN$  (1.5 mL) at  $-15$   $^{\circ}C$ . The mixture was gradually warmed over a period of 4.5 h to room temperature, quenched with  $Et_3N$ , diluted with EtOAc, and filtered through a Celite bed. The filtrate was washed successively with aq.  $NaHCO_3$ , brine, dried ( $MgSO_4$ ) and concentrated in vacuo. The residue was purified by successive chromatography, firstly on Bio-Beads SX-3 with toluene and then on  $SiO_2$  with 5 : 1  $CHCl_3$ -acetone to afford **20** (138 mg, 65%):  $[\alpha]_D - 16.2^{\circ}$  ( $c$  1.0);  $R_f$  0.43 (3 : 1  $CHCl_3$ -acetone);  $^1H$  NMR ( $CDCl_3$ )  $\delta$  7.617 (m, 4H, Phth), 5.223 (dd, 1H,  $J = 2.6, 8.8$  Hz, H-7e), 5.152 (d, 1H,  $J = 8.4$  Hz, H-1c), 5.037 (d, 1H,  $J = 3.7$  Hz, H-1f), 4.954 (d, 1H,  $J = 4.0$  Hz, H-4d), 3.758 (s, 3H, OMe), 2.084, 2.030, 2.006, 1.969, 1.874, 1.804, 1.677 (7s, 21H, 7Ac), 1.086 (s, 9H, *tert*-Bu).

Anal. Calcd for  $C_{144}H_{160}N_2O_{41}$ : C, 67.17; H, 6.26; N, 1.09. Found: C, 66.88; H, 6.27; N, 1.15.

**O - (Methyl 5 - Acetamido - 4, 7, 8, 9 - tetra - O - acetyl - 3, 5 - dideoxy - D - glycerol -  $\alpha$  - D - galacto - 2 - nonulopyranosylate) - (2  $\rightarrow$  3) - O - (2, 4, 6 - tri - O - acetyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  3) - O - [(2, 3, 4 - tri - O - acetyl -  $\alpha$  - L - fucopyranosyl) - (1  $\rightarrow$  4)] - O - (6 - O - acetyl - 2 - deoxy - 2 - phthalimido -  $\beta$  - D - glucopyranosyl) - (1  $\rightarrow$  3) - O - (2, 4, 6 - tri - O - acetyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  4) - 1, 3, 6 - tri - O - acetyl - 2 - O - pivaloyl - D - glucopyranose (21).** A mixture of **20** (119 mg, 46  $\mu$ mol) and  $Pd(OH)_2 \cdot C$  (119 mg) in 4 : 1 MeOH- $H_2O$  (8 mL) was stirred under  $H_2$  for 19 h at room temperature, diluted with 4 : 1 MeOH- $H_2O$  and filtered through a Celite

bed. The filtrate was coevaporated with toluene. To a solution of the residue in pyridine (3 mL) was added acetic anhydride (3 mL) and DMAP. The mixture was stirred for 5 days at room temperature and then coevaporated with toluene in vacuo. Chromatography of the residue on SiO<sub>2</sub> with 20 : 1 CHCl<sub>3</sub>-MeOH afforded **21** (75 mg, 99%) as a 1 : 1 mixture of  $\alpha$  and  $\beta$  anomers: R<sub>f</sub> 0.27 (20 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.795 (m, 4H, Phth), 6.237 (d, 0.5H, J = 3.7 Hz, H-1a( $\alpha$ )), 5.613 (d, 0.5H, J = 8.4 Hz, H-1a( $\beta$ )), 1.274 (d, 3H, J = 6.2 Hz, H-6f), 1.091, 1.104 (2s, 9H, *tert*-Bu).

Anal. Calcd for C<sub>89</sub>H<sub>116</sub>N<sub>2</sub>O<sub>52</sub> · 2H<sub>2</sub>O: C, 51.34; H, 5.81; N, 1.35. Found: C, 51.06; H, 5.50; N, 1.41.

**O** - (Methyl 5 - Acetamido - 4, 7, 8, 9 - tetra - *O* - acetyl - 3, 5 - dideoxy - D - glycerol -  $\alpha$  - D - galactol - 2 - nonulopyranosylate) - (2  $\rightarrow$  3) - *O* - (2, 4, 6 - tri - *O* - acetyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  3) - *O* - [(2, 3, 4 - tri - *O* - acetyl -  $\alpha$  - L - fucopyranosyl) - (1  $\rightarrow$  4)] - *O* - (6 - *O* - acetyl - 2 - deoxy - 2 - phthalimido -  $\beta$  - D - glucopyranosyl) - (1  $\rightarrow$  3) - *O* - (2, 4, 6 - tri - *O* - acetyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  4) - 3, 6 - di - *O* - acetyl - 2 - *O* - pivaloyl - D - glucopyranose (**22**). A mixture of **21** (98 mg, 48  $\mu$ mol) and H<sub>2</sub>NNH<sub>2</sub>-AcOH (9 mg, 96  $\mu$ mol) in DMF (1 mL) was stirred for 1 h at room temperature. The mixture was diluted with EtOAc and washed with brine. The organic layer was dried (MgSO<sub>4</sub>), and concentrated in vacuo. Chromatography of the residue on Sephadex LH-20 with 1 : 1 CHCl<sub>3</sub>-MeOH afforded **22** (90 mg, 94%): R<sub>f</sub> 0.44 (10 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  3.783 (s, 3H, OMe), 2.18-1.81 (br, 54H, 18Ac), 1.149 (s, 9H, *tert*-Bu).

**O** - (Methyl 5 - Acetamido - 4, 7, 8, 9 - tetra - *O* - acetyl - 3, 5 - dideoxy - D - glycerol -  $\alpha$  - D - galactol - 2 - nonulopyranosylate) - (2  $\rightarrow$  3) - *O* - (2, 4, 6 - tri - *O* - acetyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  3) - *O* - [(2, 3, 4 - tri - *O* - acetyl -  $\alpha$  - L - fucopyranosyl) - (1  $\rightarrow$  4)] - *O* - (6 - *O* - acetyl - 2 - deoxy - 2 - phthalimido -  $\beta$  - D - glucopyranosyl) - (1  $\rightarrow$  3) - *O* - (2, 4, 6 - tri - *O* - acetyl -  $\beta$  - D - galactopyranosyl) - (1  $\rightarrow$  4) - 3, 6 - di - *O* - acetyl - 2 - *O* - pivaloyl -  $\alpha$  - D - glucopyranosyl Trichloroacetimidate (**23**). A solution of **22** (90 mg, 45  $\mu$ mol), CCl<sub>3</sub>CN (70  $\mu$ L, 676  $\mu$ mol), and DBU (15  $\mu$ L, 95  $\mu$ mol), in (ClCH<sub>2</sub>)<sub>2</sub> (1 mL) was stirred for 45 min at 0 °C. The reaction mixture was directly chromatographed on SiO<sub>2</sub> with 30 : 1 CHCl<sub>3</sub>-MeOH to afford **23** (89 mg, 91%): [ $\alpha$ ]<sub>D</sub> -3.8 ° (c 1.0); R<sub>f</sub> 0.45 (15 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.613 (s, 1H, C = NH), 7.783 (m, 4H, Phth), 6.452 (d, 1H, J = 3.7 Hz, H-1a), 5.245 (d, 1H, J = 2.6 Hz, H-4b), 5.217 (d, 1H, J = 3.3 Hz, H-4d), 5.200 (d, 1H, J = 3.7 Hz, H-1f),

5.098 (d, 1H,  $J = 10.3$  Hz, NH), 5.001 (d, 1H,  $J = 7.3$  Hz, H-1d), 4.754 (t, 1H,  $J = 9.6$  Hz, H-2d), 4.658 (dd,  $J = 8.1, 9.9$  Hz, H-2b), 4.470 (d, 1H,  $J = 8.1$  Hz, H-1c), 4.229 (d, 1H,  $J = 8.1$  Hz, H-1b), 3.782 (s, 3H, OMe), 2.462 (dd, 1H,  $J = 4.8, 12.5$  Hz, H-3e<sub>eq</sub>), 2.166, 2.160, 2.118, 2.103, 2.093, 2.079, 2.077, 2.074, 2.047, 2.001, 1.967, 1.842, 1.832, 1.810, (14s, 51H, 17Ac), 1.274 (d, 3H,  $J = 6.6$  Hz, H-6f), 1.099 (s, 9H, *tert*-Bu).

Anal. Calcd for  $C_{89}H_{114}Cl_3N_3O_{51}$ : C, 49.76; H, 5.35; N, 1.96. Found: C, 49.98; H, 5.27; N, 2.01.

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-acetyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-*O*-(6-*O*-acetyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-(3,6-di-*O*-acetyl-2-*O*-pivaloyl- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 1)-(2*S*, 3*R*, 4*E*)-2-azido-3-*O*-*tert*-butyldiphenylsilyl-4-octadecene-1,3-diol (24).** To a stirred mixture of **23** (49 mg, 23  $\mu$ mol) and (2*S*,3*R*,4*E*)-2-azido-3-*O*-*tert*-butyldiphenylsilyl-4-octadecene-1,3-diol (**3**, 26 mg, 46  $\mu$ mol) and 4A molecular sieves (500 mg) in  $CHCl_3$  (1 mL) was added  $BF_3 \cdot Et_2O$  (4  $\mu$ L, 46  $\mu$ mol) at  $-15$   $^\circ C$ . The mixture was stirred for 30 minutes and gradually warmed to room temperature, diluted with  $CHCl_3$ , and filtered through a Celite bed. The filtrate was washed successively with aq.  $NaHCO_3$ , brine, dried ( $MgSO_4$ ) and concentrated in vacuo. Chromatography of the residue over  $SiO_2$  with 2 : 1 toluene-acetone afforded **24** (50 mg, 86%):  $[\alpha]_D^{25} -25.7^\circ$  ( $c$  0.8);  $R_f$  0.61 (1 : 1 toluene-acetone);  $^1H$  NMR ( $CDCl_3$ )  $\delta$  5.262 (d, 1H,  $J = 2.6$  Hz, H-4b), 5.010 (d, 1H,  $J = 8.4$  Hz, H-1d), 4.809 (dd, 1H,  $J = 7.7, 9.5$  Hz, H-2d), 4.630 (dd, 1H,  $J = 8.1, 9.9$  Hz, H-2b), 4.467 (d, 1H,  $J = 7.7$  Hz, H-1c), 4.282 (d, 1H,  $J = 8.1$  Hz, H-1b), 4.172 (d, 1H,  $J = 8.1$  Hz, H-1a), 3.783 (s, 3H, OMe), 2.461 (dd, 1H,  $J = 4.8, 12.8$  Hz, H-3e<sub>eq</sub>), 2.161, 2.159, 2.105, 2.099, 2.079, 2.072, 2.060, 2.047, 1.999, 1.967, 1.957, 1.941, 1.961, 1.824, 1.810 (15s, 51H, 17Ac), 1.067 (s, 9H, *tert*-Bu), 0.883 (t, 3H,  $J = 7.0$  Hz,  $CH_2Me$ ).

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-acetyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-*O*-(6-*O*-acetyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-(3,6-di-*O*-acetyl-2-*O*-pivaloyl- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 1)-(2*S*, 3*R*, 4*E*)-2-azido-3-*O*-*tert*-butyldiphenylsilyl-4-octadecene-1,3-diol (24).**



pyranosyl) - (1→3) - *O* - (2,4,6 - tri-*O* - acetyl - β - D - galactopyranosyl) - (1→4) - (3,6 - di-*O* - acetyl-2-*O* - pivaloyl - β - D - glucopyranosyl) - (1→1) - (2*S*, 3*R*, 4*E*) - 3-*O* - *tert*-butyldiphenylsilyl- 2 - tetracosanoylamino- 4 - octadecene-1,3 - diol (**26**). To a solution of **24** (35 mg, 14 μmol) in toluene (2 mL) and H<sub>2</sub>O (1 mL) were added triphenylphosphine (7 mg, 28 μmol). After heating under reflux for 19 h, the solvent was evaporated in vacuo, the residue was applied to a column of Sephadex LH-20 and was eluted with 1 : 1 MeOH-CHCl<sub>3</sub> to afford the amino derivative **25** (28 mg, 82%). A mixture of thus obtained **25** (28 mg, 11 μmol), tetracosanoic acid (9 mg, 262 μmol), 2-chloro-1-methylpyridinium iodide (7 mg, 26 μmol) and Bu<sub>3</sub>N (11 μL, 52 μmol) in (ClCH<sub>2</sub>)<sub>2</sub> (1 mL) was stirred for 1 h at room temperature. The mixture was diluted with AcOEt, washed with aq. NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. The residue was purified by successive chromatography on Sephadex LH-20 with 1 : 1 MeOH-CHCl<sub>3</sub> and on SiO<sub>2</sub> with 3 : 1 toluene-acetone to give **26** (28 mg, 88%): [α]<sub>D</sub> -18.5° (*c* 1.0); R<sub>f</sub> 0.38 (25 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.408 (d, 1H, J = 8.8 Hz, NH), 5.013 (d, 1H, J = 8.4 Hz, H-1d), 4.816 (dd, 1H, J = 8.1, 9.9 Hz, H-2d), 4.629 (dd, 1H, J = 8.1, 9.5 Hz, H-2b), 4.471 (d, 1H, J = 8.1 Hz, H-1c), 4.337 (d, 1H, J = 7.7 Hz, H-1b), 3.783 (s, 3H, OMe), 2.463 (dd, 1H, J = 4.4, 12.8 Hz, H-3<sub>eeq</sub>), 2.164, 2.160, 2.115, 2.106, 2.080, 2.074, 2.050, 2.002, 1.967, 1.942, 1.929, 1.830, 1.811 (13s, 51H, 17Ac), 1.092 (s, 9H, *tert*-Bu), 0.880 (t, 6H, J = 7.0 Hz, 2CH<sub>2</sub>Me).

*O* - (Lithium 5 - Acetamido - 4,7,8,9 - tetra-*O* - acetyl - 3,5 - dideoxy - D - glycerol - α - D - galactopyranosylate) - (2→3) - *O* - (2,4,6 - tri-*O* - acetyl - β - D - galactopyranosyl) - (1→3) - *O* - [(2,3,4 - tri-*O* - acetyl - α - L - fucopyranosyl) - (1→4)] - *O* - (6-*O* - acetyl - 2 - deoxy - 2 - phthalimido - β - D - glucopyranosyl) - (1→3) - *O* - (2,4,6 - tri-*O* - acetyl - β - D - galactopyranosyl) - (1→4) - 3,6 - di-*O* - acetyl-2-*O* - pivaloyl - β - D - glucopyranosyl) - (1→1) - (2*S*, 3*R*, 4*E*) - 3-*O* - *tert*-butyldiphenylsilyl - 2 - tetracosanoylamino- 4 - octadecene-1,3 - diol (**27**). A solution of **26** (28 mg, 10 μmol) in pyridine (1.5 mL) was added dropwise onto LiI (30 mg, 226 μmol, dried at 200 °C for 12 h in vacuo), and the mixture was heated for 12.5 h at reflux under argon. The reaction mixture was chromatographed over Sephadex LX-20 with 1 : 1 CHCl<sub>3</sub>-MeOH to afford **27** (22 mg, 79%): R<sub>f</sub> 0.65 (5 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CD<sub>3</sub>OD) δ 5.075 (d, 1H, J = 8.4 Hz, H-1d), 4.815 (dd, 1H, J = 8.1, 9.5 Hz, H-2d), 4.773 (t, 1H, J = 9.2 Hz, H-2a), 4.641 (dd, 1H, J = 8.1, 9.9 Hz, H-2b), 4.548 (d, 1H, J = 8.1 Hz, H-1c), 2.486 (dd, 1H, J = 4.3, 12.8 Hz, H-3<sub>eeq</sub>),

2.185, 2.181, 2.134, 2.129, 2.110, 2.105, 2.072, 2.063, 2.028, 1.966, 1.961, 1.956, 1.823, 1.801 (14s, 51H, 17Ac), 1.099 (s, 9H, *tert*-Bu), 0.891 (t, 6H,  $J = 7.0$  Hz, 2CH<sub>2</sub>Me).

***O*-(Sodium 5-Acetamido-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*- $\beta$ -D-galactopyranosyl-(1 $\rightarrow$ 3)-*O*-[ $\alpha$ -L-fucopyranosyl-(1 $\rightarrow$ 4)]-*O*-(2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*- $\beta$ -D-galactopyranosyl-(1 $\rightarrow$ 4)-*O*- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 1)-(2*S*, 3*R*, 4*E*)-2-tetracosanoylamino-4-octadecene-1,3-diol. (1) (Sialyl monomeric Le<sup>a</sup> ganglioside)** To a solution of **27** (22 mg, 7.8  $\mu$ mol) in EtOH (5 mL) was added MeNHNH<sub>2</sub> (2.5 mL), and the mixture was stirred for 18 h at 80 °C, and then concentrated in vacuo. The residue was purified over Sephadex LH-20 with MeOH to afford the amino derivative. To the solution of amino derivative in MeOH (2 mL) was added acetic anhydride (70  $\mu$ L), and the mixture was stirred for 1 h at room temperature and concentrated in vacuo. The residue in 1 : 1 MeOH-THF (0.6 mL) was treated with 1N-NaOH (0.3 mL) for 1 day at room temperature, and the mixture was chromatographed on a sephadex LH-20 with 60 : 30 : 5 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O and then concentrated. THF (200  $\mu$ L) and Bu<sub>4</sub>NF (528  $\mu$ L) were added to the residue, and the mixture was stirred for 14 h at 60 °C and then concentrated in vacuo. The residue was purified over Sephadex LH-20 with 60 : 30 : 5 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O to give **1** (8 mg, 59%): R<sub>f</sub> 0.25 (60 : 30 : 5 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O); <sup>1</sup>H NMR (50 : 1 (CD<sub>3</sub>)<sub>2</sub>SO-D<sub>2</sub>O)  $\delta$  5.562 (dt, 1H,  $J = 7.0, 15.0$  Hz, H-5Cer), 5.366 (dd, 1H,  $J = 7.0, 15.4$  Hz, H-4Cer), 4.813 (d, 1H,  $J = 3.7$  Hz, H-1f), 4.771 (d, 1H,  $J = 8.1$  Hz, H-1c), 4.366 (d, 1H,  $J = 7.7$  Hz, H-1d), 4.289 (d, 1H,  $J = 6.6$  Hz, H-1b), 4.185 (d, 1H,  $J = 8.1$  Hz, H-1a), 2.773 (dd, 1H,  $J = 4.8, 12.1$  Hz, H-3<sub>eeq</sub>), 1.850, 1.899 (2s, 6H, 2NAc), 1.023 (d, 3H,  $J = 6.6$  Hz, H-6f), 0.853 (t, 6H,  $J = 7.0$  Hz, 2CH<sub>2</sub>Me); FABMS (TEA matrix):  $m/z$  (M-Na)<sup>-</sup> 1775.

**Allyl *O*-(Methyl-5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(6-*O*-benzyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-benzyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-6-*O*-benzyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranoside (30).** To a stirred mixture of 1 : 1 HgBr<sub>2</sub>-Hg(CN)<sub>2</sub> (161 mg), and 1 : 2 mixture of 3A-4A molecular sieves (3.6 g) was added a solution of **28** (466 mg, 0.42 mmol) in CH<sub>3</sub>CN (2 mL), and the mixture was stirred for 1 h at room temperature. A solution of **29** (661 mg, 1.26 mmol) in CH<sub>3</sub>CN (2 mL) was added, and this was stirred for 1 day at room

temperature. The mixture was quenched with  $\text{Et}_3\text{N}$ , diluted with  $\text{EtOAc}$ , and filtered through a Celite bed. The filtrate was washed with aq.  $\text{NaHCO}_3$ , brine, dried ( $\text{MgSO}_4$ ) and concentrated in vacuo. The residue was purified by successive chromatography, first on Bio-Beads SX-3 with toluene and then on  $\text{SiO}_2$  with 1 : 2 toluene- $\text{EtOAc}$ , to give **30** (472 mg, 71%):  $[\alpha]_D^{25} +25.6^\circ$  (*c* 1.0);  $R_f$  0.25 (1 : 2 toluene- $\text{AcOEt}$ );  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.666 (m, 1H,  $-\text{CH}_2-\text{CH}=\text{CH}_2$ ), 5.120 (d, 1H,  $J = 3.7$  Hz, H-1i), 4.348 (dd, 1H,  $J = 8.4, 10.6$  Hz, H-2e), 3.739 (s, 3H, OMe), 2.425 (dd, 1H,  $J = 4.8, 13.2$  Hz, H-3geq), 2.049, 2.044, 2.027, 1.986, 1.872 (5s, 15H, 5Ac), 1.124 (d, 3H,  $J = 6.6$  Hz, H-6i).

Anal. Calcd for  $\text{C}_{84}\text{H}_{96}\text{N}_2\text{O}_{28}$ : C, 63.79; H, 6.12; N, 1.77. Found: C, 63.68; H, 6.10; N, 1.74.

**Allyl O-(Methyl-5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-O-(2-O-acetyl-6-O-benzyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-O-[(2,3,4-tri-O-benzyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-6-O-benzyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranoside (31).** To a solution of **30** (507 mg, 0.321 mmol) in pyridine (15 mL) was added acetic anhydride (15 mL), and the mixture was stirred for 1 day at room temperature, and coevaporated with toluene. Chromatography of the residue over  $\text{SiO}_2$  with 3 : 1 toluene-acetone afforded **31** (490 mg, 94%):  $[\alpha]_D -28.8^\circ$  (*c* 1.0);  $R_f$  0.39 (5 : 1 toluene-MeOH);  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.593 (m, 1H,  $-\text{CH}_2-\text{CH}=\text{CH}_2$ ), 5.119 (d, 1H,  $J = 3.3$  Hz, H-1i), 4.861 (t, 1H,  $J = 9.5$  Hz, H-2f), 4.378 (d, 1H,  $J = 7.7$  Hz, H-1f), 4.288 (t, 1H,  $J = 10.6$  Hz, H-2e), 3.969 (t, 1H,  $J = 9.5$  Hz, H-3e), 3.664 (s, 3H, OMe), 2.519 (dd, 1H,  $J = 4.4, 12.8$  Hz, H-3geq), 2.091, 2.044, 2.031, 1.993, 1.975, 1.824 (6s, 18H, 6Ac), 1.740 (t, 1H,  $J = 12.8$  Hz, H-3gax), 1.291 (d, 3H,  $J = 6.6$  Hz, H-6i).

Anal. Calcd for  $\text{C}_{88}\text{H}_{100}\text{N}_2\text{O}_{30} \cdot \text{H}_2\text{O}$ : C, 63.45; H, 6.05; N, 1.68. Found: C, 63.55; H, 6.05; N, 1.62.

**O-(Methyl-5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-O-(2-O-acetyl-6-O-benzyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-O-[(2,3,4-tri-O-benzyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-6-O-benzyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranose (32).** To a solution of (1,5-cyclooctadiene)bis(methylphenylphosphine) iridium hexafluoro-phosphate (27 mg, 22.7  $\mu\text{mol}$ ), which was activated by  $\text{H}_2$  in THF (5 mL), was added a solution of **31** (361 mg, 227  $\mu\text{mol}$ ) in THF (5 mL). The mixture was stirred for 4 h at room temperature, then  $\text{I}_2$  (341 mg, 1.34 mmol) and  $\text{H}_2\text{O}$  (6 mL) were

added. Stirring was continued for 0.5 h at room temperature. The mixture was diluted with  $\text{CHCl}_3$ , washed successively with aq. sodium thiosulfate, aq.  $\text{NaHCO}_3$ , brine, dried ( $\text{MgSO}_4$ ) and concentrated in vacuo. The residue was chromatographed on  $\text{SiO}_2$  with 2 : 1  $\text{CHCl}_3$ -acetone to afford **32** (360 mg, qu.):  $[\alpha]_D -24.8^\circ$  (*c* 1.0);  $R_f$  0.17 (3 : 1  $\text{CHCl}_3$ -acetone);  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.276 (dd, 1H,  $J = 1.8, 8.8$  Hz, H-7g), 5.141 (t, 1H,  $J = 8.8$  Hz, H-2f), 5.050 (d, 1H,  $J = 3.3$  Hz, H-1i), 4.147 (dd, 1H,  $J = 8.4, 10.6$  Hz, H-2e), 3.968 (t, 1H,  $J = 9.5$  Hz, H-3e), 3.814 (t, 1H,  $J = 8.8$  Hz, H-4e), 3.660 (s, 3H, OMe), 2.516 (dd, 1H,  $J = 4.4, 12.8$  Hz, H-3geg), 2.056, 2.025, 2.002, 1.962, 1.840 (5s, 18H, 6Ac).

Anal. Calcd for  $\text{C}_{83}\text{H}_{94}\text{N}_2\text{O}_{29}$ : C, 62.95; H, 5.98; N, 1.77. Found: C, 62.85; H, 6.05; N, 1.75.

**O**- (Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate) - (2 $\rightarrow$ 3)-*O*- (2-*O*-acetyl-6-*O*-benzyl- $\beta$ -D-galactopyranosyl) - (1 $\rightarrow$ 3)-*O*- [(2,3,4-tri-*O*-benzyl- $\alpha$ -L-fucopyranosyl) - (1 $\rightarrow$ 4)] - 6-*O*-benzyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl Trichloroacetimidate (**33**). A mixture of **32** (49 mg, 31  $\mu\text{mol}$ ),  $\text{CCl}_3\text{CN}$  (46  $\mu\text{L}$ , 0.46 mmol), and DBU (3  $\mu\text{L}$ , 22  $\mu\text{mol}$ ) in  $\text{CH}_2\text{Cl}_2$  (1.0 mL) was stirred for 1 h at  $0^\circ\text{C}$ . The reaction mixture was directly chromatographed on  $\text{SiO}_2$  with 3 : 1  $\text{CHCl}_3$ -acetone to afford **33** (47 mg, 89%):  $[\alpha]_D -17.5^\circ$  (*c* 1.0);  $R_f$  0.32 (3 : 1  $\text{CHCl}_3$ -acetone);  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  8.459 (s, 1H, C = NH), 6.215 (d, 1H,  $J = 8.8$  Hz, H-1e), 5.138 (d, 1H,  $J = 3.7$  Hz, H-1i), 4.927 (1H,  $J = 10.9$  Hz, H-2f), 4.562 (t, 1H,  $J = 10.6$  Hz, H-2e), 4.396 (d, 1H,  $J = 7.7$  Hz, H-1f), 4.086 (t, 1H,  $J = 9.5$  Hz, H-3e), 3.659 (s, 3H, OMe), 3.540 (br-d, 1H, H-4f), 2.525 (dd, 1H,  $J = 4.4, 13.2$  Hz, H-3geg), 2.091, 2.048, 2.026, 2.003, 1.954, 1.840 (6s, 18H, 6Ac), 1.288 (d, 3H,  $J = 6.2$  Hz, H-6i).

**Methyl O**- (Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate) - (2 $\rightarrow$ 3)-*O*- (2-*O*-acetyl-6-*O*-benzyl- $\beta$ -D-galactopyranosyl) - (1 $\rightarrow$ 3)-*O*- [(2,3,4-tri-*O*-benzyl- $\alpha$ -L-fucopyranosyl) - (1 $\rightarrow$ 4)] - 6-*O*-benzyl-2-deoxy-2-phthalimido-1-thio- $\beta$ -D-glucopyranoside (**34**). To a stirred mixture of **33** (233 mg, 135  $\mu\text{mol}$ ) and powdered molecular sieves (4A, 0.3 g) in dry  $\text{CH}_2\text{Cl}_2$  (1.5 mL) was added  $\text{Bu}_3\text{SnSMe}$  (69  $\mu\text{L}$ , 0.20 mmol) at  $-10^\circ\text{C}$ . After stirring for 10 min at  $-10^\circ\text{C}$ , boron trifluoride etherate (24  $\mu\text{L}$ , 18  $\mu\text{mol}$ ) was added, and the mixture was stirred for 2 h. The mixture was diluted with EtOAc, filtered through Celite and the filtrate was washed successively with aq. KF, aq.

NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. Chromatography of the residue on SiO<sub>2</sub> with 6 : 1 CHCl<sub>3</sub>-acetone and gel filtration over Bio-Beads S-X1 with toluene afforded **34** (209 mg, 96%): [ $\alpha$ ]<sub>D</sub> +25.6° (*c* 1.0); R<sub>f</sub> 0.32 (3 : 1 CHCl<sub>3</sub>-acetone); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.139 (d, 1H, *J* = 3.7 Hz, H-1i), 4.364 (d, 1H, *J* = 7.3 Hz, H-1f), 3.973 (t, 1H, *J* = 9.2 Hz, H-3e), 3.826 (t, 1H, *J* = 8.8 Hz, H-4e), 3.555 (br-d, 1H, H-4f), 2.514 (dd, 1H, *J* = 4.8, 12.5 Hz, H-3geq), 2.093, 2.080, 2.051, 2.036, 2.003, 1.970, 1.838 (7s, 21H, 6Ac and 1SMe), 1.740 (t, 1H, *J* = 12.5 Hz, H-3gax), 1.284 (d, 3H, *J* = 6.6 Hz, H-6i).

Anal. Calcd for C<sub>84</sub>H<sub>96</sub>N<sub>2</sub>O<sub>28</sub>S<sub>1</sub> · 2H<sub>2</sub>O: C, 61.16; H, 6.11; N, 1.70. Found: C, 61.01; H, 6.13; N, 1.74.

**Benzyl O-(Methyl-5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-O-(2,4-di-O-acetyl-6-O-benzyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-O-[(2,3,4-tri-O-benzyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-O-(6-O-benzyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-O-(6-O-benzyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-O-[(2,3,4-tri-O-benzyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-O-(2-acetamido-6-O-benzyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-O-(2,4,6-tri-O-benzyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-3,6-di-O-benzyl-2-O-pivaloyl- $\beta$ -D-glucopyranoside (**36**).** (Method A) To a stirred mixture of **35** (27 mg, 14  $\mu$ mol) and powdered molecular sieves (3A, 0.3 g) in dry CH<sub>3</sub>CN (3 mL) was added a solution of **33** (39 mg, 22  $\mu$ mol) in dry CH<sub>3</sub>CN (1.5 mL) at -40 °C. After stirring for 10 min at -40 °C, boron trifluoride etherate (2.9  $\mu$ L, 44  $\mu$ mol) was added, and the mixture was stirred for 2 h. The mixture was diluted with EtOAc, filtered through Celite and the filtrate was washed successively with aq. NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. Chromatography of the residue on SiO<sub>2</sub> with 6 : 1 CHCl<sub>3</sub>-acetone and gel filtration over Bio-Beads SX-1 with toluene afforded **36** (8 mg, 15%). (Method B) To a stirred mixture of **35** (40 mg, 20  $\mu$ mol) and powdered molecular sieves (3A, 0.3 g) in dry CH<sub>3</sub>CN (3 mL) was added a solution of **34** (23 mg, 14  $\mu$ mol) in dry CH<sub>3</sub>CN (1.5 mL) at -20 °C. After stirring for 10 min at -10 °C, AgOTf (13 mg, 55  $\mu$ mol) and PhSeCl (10 mg, 55  $\mu$ mol) were added, and the mixture was stirred for 2 h, and worked up as described in Method A. Chromatography of the residue on SiO<sub>2</sub> with 6 : 1 CHCl<sub>3</sub>-acetone and gel filtration over Bio-Beads SX-1 with toluene afforded **36** (23 mg, 47%): [ $\alpha$ ]<sub>D</sub> -25.8° (*c* 1.0); R<sub>f</sub> 0.53 (3 : 1 CHCl<sub>3</sub>-acetone); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  5.234 (d, 1H, *J* = 7.7 Hz, H-1e), 3.662 (s,

3H, OMe), 1.279 (d, 3H,  $J = 6.2$  Hz, H-6h), 1.121 (s, 9H, *tert*-Bu), 0.798 (d, 3H,  $J = 6.2$  Hz, H-6i).

Anal. Calcd for  $C_{197}H_{221}N_3O_{54} \cdot 3H_2O$ : C, 66.70; H, 6.45; N, 1.18. Found: C, 66.40; H, 6.23; N, 1.18.

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-acetyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-*O*-(6-*O*-acetyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-acetyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-*O*-(2-acetamido-6-*O*-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-1,3,6-tri-*O*-acetyl-2-*O*-pivaloyl- $\beta$ -D-glucopyranose (37).** A mixture of **36** (81 mg, 23  $\mu$ mol) and 20% Pd(OH)<sub>2</sub>-C (81 mg) in 4 : 1 MeOH-H<sub>2</sub>O (8 mL) was stirred under H<sub>2</sub> for 19 h at room temperature, diluted with 4 : 1 MeOH-H<sub>2</sub>O and filtered through a Celite bed. The filtrate was coevaporated with toluene. To a solution of the residue in pyridine (3 mL) was added acetic anhydride (2 mL) and a catalytic amount of DMAP. The mixture was stirred for 1 day at room temperature and then coevaporated with toluene in vacuo. Chromatography of the residue on sephadex LH-20 with 1 : 1 CHCl<sub>3</sub>-MeOH afforded **37** (56 mg, 85%) as a 1 : 1 mixture of  $\alpha$  and  $\beta$  anomers:  $R_f$  0.21 (20 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  6.289 (d, 0.5H,  $J = 4.0$  Hz, H-1a( $\alpha$ )), 5.692 (d, 0.5H,  $J = 8.1$  Hz, H-1a( $\beta$ )), 3.785 (s, 3H, OMe), 1.272 (d, 3H,  $J = 6.2$  Hz, H-6h), 1.134, 1.121 (2s, 9H, *tert*-Bu), 0.830 (d, 3H,  $J = 6.2$  Hz, H-6i).

Anal. Calcd for  $C_{123}H_{163}N_3O_{73} \cdot H_2O$ : C, 51.48; H, 5.80; N, 1.46. Found: C, 51.41; H, 5.44; N, 1.48.

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-acetyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-*O*-(6-*O*-acetyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-*O*-[(2,3,4-tri-*O*-acetyl- $\alpha$ -L-fucopyranosyl)-(1 $\rightarrow$ 4)]-*O*-(2-acetamido-6-*O*-acetyl-2-deoxy- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-acetyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-3,6-di-*O*-acetyl-2-*O*-pivaloyl- $\beta$ -D-glucopyranose (38).** A mixture of **37** (56 mg, 20  $\mu$ mol) and H<sub>2</sub>NNH<sub>2</sub>-AcOH (5 mg, 60  $\mu$ mol) in DMF (1 mL)

was stirred for 1.5 h at room temperature. The mixture was chromatographed on Sephadex LH-20 with MeOH to afford **38** (52 mg, 94%):  $R_f$  0.14 (20 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 3.784 (s, 3H, OMe), 1.133, 1.120 (2s, 9H, *tert*-Bu).

Anal. Calcd for C<sub>121</sub>H<sub>161</sub>N<sub>3</sub>O<sub>72</sub> · H<sub>2</sub>O: C, 51.40; H, 5.74; N, 1.49. Found: C, 51.28; H, 5.79; N, 1.61.

**O - (Methyl 5 -Acetamido- 4,7,8,9 -tetra-O-acetyl- 3,5 -dideoxy-D-glycero- α -D-galacto - 2 -nonulopyranosylonate) - (2→3) -O - (2, 4, 6 - tri -O -acetyl- β -D - galactopyranosyl) - (1→3) -O - [(2,3,4 -tri-O -acetyl- α - L - fucopyranosyl) - (1→4)] -O - (6-O-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl) - (1→3) -O - (2, 4, 6 - tri -O -acetyl- β -D - galactopyranosyl) - (1→3) -O - [(2, 3, 4 -tri-O -acetyl-α-L-fucopyranosyl) - (1→4)] -O - (2-acetamido-6-O-acetyl-2-deoxy- β -D - glucopyranosyl) - (1→3) -O - (2, 4, 6 - tri -O -acetyl- β -D - galactopyranosyl) - (1→4) - 3,6 - di -O -acetyl-2-O -pivaloyl-α-D-glucopyranosyl Trichloroacetimidate (39).** A solution of **38** (52 mg, 18 μmol), CCl<sub>3</sub>CN (29 μL, 260 μmol), and DBU (6 μL, 38 μmol) in (ClCH<sub>2</sub>)<sub>2</sub> (1 mL) was stirred for 2 h at 0 °C. The reaction mixture was directly chromatographed on SiO<sub>2</sub> with 35 : 1 CHCl<sub>3</sub>-MeOH to afford **39** (34 mg, 62%):  $[\alpha]_D$  -24.5° (c 1.0);  $R_f$  0.61 (20 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.660 (s, 1H, C = NH), 6.505 (d, 1H, J = 3.6 Hz, H-1a), 5.568 (t, 1H, J = 9.5 Hz, H-3a), 5.359 (br-s, 1H, H-1h or 1i), 5.307 (d, 1H, J = 4.0 Hz, H-4b or 4d), 5.269 (d, 2H, J = 3.3 Hz, H-4d or 4b and H-1i or 1h), 5.181 (br-s, 1H, H-4h or 4i), 5.047 (d, 1H, J = 9.3 Hz, H-1e), 5.007 (br-s, 1H, H-4i or 4h), 4.975 (d, 1H, J = 8.1 Hz, H-1c), 4.955 (d, 1H, J = 4.0 Hz, H-4f), 4.927 (t, 1H, J = 9.2 Hz, H-2d or 2b), 4.814 (t, 1H, J = 9.9 Hz, H-2c or 2f), 4.595 (d, 1H, J = 8.0 Hz, H-1f), 4.479 (t, 1H, J = 9.5 Hz, H-2b or 2d), 4.377 (d, 1H, J = 8.4 Hz, H-1b or 1d), 4.361 (d, 1H, J = 7.7 Hz, H-1d or 1b), 4.298 (br-dd, 1H, H-3f), 3.785 (s, 3H, OMe), 1.272 (d, 3H, J = 6.2 Hz, H-6h), 1.130 (s, 9H, *tert*-Bu), 0.832 (d, 3H, J = 6.6 Hz, H-6i).

**O - (Methyl 5 -Acetamido- 4,7,8,9 -tetra-O-acetyl- 3,5 -dideoxy-D-glycero- α -D-galacto - 2 - nonulopyranosylonate) - (2→3) -O - (2, 4, 6 - tri -O -acetyl- β -D - galactopyranosyl) - (1→3) -O - [(2,3,4 -tri-O -acetyl- α - L - fucopyranosyl) - (1→4)] -O - (6-O-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl) - (1→3) -O - (2, 4, 6 - tri -O -acetyl-β-D-galactopyranosyl) - (1→3) -O - [(2, 3, 4 -tri-O -acetyl-α-L-fucopyranosyl) - (1→4)] -O - (2-acetamido-6-O-acetyl-2-deoxy- β -D - glucopyranosyl) - (1→3) -O - (2, 4, 6 - tri -O -acetyl- β -D - galacto-**

**pyranosyl)-(1→4)-3,6-di-*O*-acetyl-2-*O*-pivaloyl-β-D-glucopyranosyl Fluoride (40).** A mixture of **38** (10 mg, 4 μmol) and DAST (2 μL, 14 μmol) in (CICH<sub>2</sub>)<sub>2</sub> (1 mL) was stirred for 2 h at -15 °C. The mixture was diluted with EtOAc, filtered through Celite and the filtrate was washed successively with aq. NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. Chromatography of the residue on SiO<sub>2</sub> with 35 : 1 CHCl<sub>3</sub>-MeOH afforded **40** (9 mg, qu.): R<sub>f</sub> 0.24 (20 : 1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.353 (br-s, H-1h or H-1i), 5.304 (br-s, H-4b or H-4d), 5.181 (br-s, H-4h or H-4i), 4.502 (d, 1H, J = 9.0 Hz, H-1f), 4.471 (t, 1H, J = 9.2 Hz, H-2d or H-2b), 4.375 (br-d, H-1b or H-1d), 4.362 (d, 1H, J = 7.3 Hz, H-1d or H-1b), 3.786 (s, 3H, OMe), 1.190 (s, 9H, *tert*-Bu).

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2→3)-*O*-(2,4,6-tri-*O*-acetyl-β-D-galactopyranosyl)-(1→3)-*O*-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→4)]-*O*-(6-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1→3)-*O*-(2,4,6-tri-*O*-acetyl-β-D-galactopyranosyl)-(1→3)-*O*-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1→4)]-*O*-(2-acetamido-6-*O*-acetyl-2-deoxy-β-D-glucopyranosyl)-(1→3)-*O*-(2,4,6-tri-*O*-acetyl-β-D-galactopyranosyl)-(1→4)-(3,6-di-*O*-acetyl-2-*O*-pivaloyl-β-D-glucopyranosyl)-(1→1)-(2*S*, 3*R*, 4*E*)-3-*O*-benzoyl-2-tetracosanoylamino-4-octadecene-1,3-diol (41).** (Method A) To a stirred mixture of **39** (34 mg, 11 μmol) and **4** (17 mg, 22 μmol) and 4A molecular sieves (500 mg) in CHCl<sub>3</sub> (1 mL) was added boron trifluoride etherate (8 μL, 44 μmol) at -10 °C. The mixture was stirred for 1 h. The mixture was quenched with Et<sub>3</sub>N, diluted with CHCl<sub>3</sub> and filtered through a Celite bed. The filtrate was washed successively with aq. NaHCO<sub>3</sub>, brine, dried (MgSO<sub>4</sub>) and concentrated in vacuo. Chromatography of the residue over SiO<sub>2</sub> with 2 : 1 toluene-acetone afforded **41** (14 mg, 34%). (Method B) To a stirred mixture of **40** (16 mg, 6 μmol), **4** (8 mg, 11 μmol) and 4A molecular sieves (500 mg) in CHCl<sub>3</sub> (1 mL) were added SnCl<sub>2</sub> (5 mg, 33 μmol) and AgOTf (17 mg, 86 μmol) at -10 °C. The mixture was stirred for 1 h, and worked up as described in Method A. Chromatography of the residue over SiO<sub>2</sub> with 2 : 1 toluene-acetone afforded **41** (8 mg, 40%): [α]<sub>D</sub> -36.9° (*c* 1.0); R<sub>f</sub> 0.44 (1 : 1 toluene-acetone); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.869 (dt, 1H, J = 7.3, 14.7 Hz, 5-Cer), 5.714 (d, 1H, J = 9.5 Hz, NH), 5.532 (t, 1H, J = 7.7 Hz, 3-Cer), 5.451 (dd, 1H, J = 7.7, 15.0 Hz, H-4Cer), 5.355 (d, 1H, J = 2.2 Hz, H-4h or 4i), 5.305 (d, 1H, J = 3.3 Hz, H-4b or



4d), 5.255 (br-s, 1H, H-4i or 4h), 5.246 (br-s, 1H, H-4d or 4b), 5.215 (br-s, 1H, H-1h or 1i), 5.497 (br-s, 1H, H-1i or 1h), 4.969 (d, 2H,  $J = 8.1$  Hz, H-1c and 1e), 4.948 (d, 1H,  $J = 2.9$  Hz, H-4f), 4.870 (t, 1H,  $J = 8.1$  Hz, H-2b or H-2d), 4.804 (t, 1H,  $J = 7.3$  Hz, H-2f), 4.504 (d, 1H,  $J = 7.7$  Hz, H-1f), 4.466 (t, 1H,  $J = 8.8$  Hz, H-2d or 2b), 4.406 (d, 1H,  $J = 8.1$  Hz, H-1a), 4.369 (d, 1H,  $J = 7.7$  Hz, H-1b or 1d), 4.262 (d, 1H,  $J = 8.1$  Hz, H-1d or 1b), 3.785 (s, 3H, OMe), 1.142 (s, 9H, *tert*-Bu), 0.849 (d, 3H,  $J = 6.2$  Hz, H-6i).

***O* - (Sodium 5-Acetamido-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylate) - (2 $\rightarrow$ 3) - *O* -  $\beta$  - D - galactopyranosyl - (1 $\rightarrow$ 3) - *O* - [ $\alpha$  - L - fucopyranosyl - (1 $\rightarrow$ 4)] - *O* - acetamido - 2-deoxy - 2 -  $\beta$  - D - glucopyranosyl - (1 $\rightarrow$ 3) - *O* -  $\beta$  - D - galactopyranosyl - (1 $\rightarrow$ 3) - *O* - [ $\alpha$  - L - fucopyranosyl - (1 $\rightarrow$ 4)] - *O* - (2-acetamido-2-deoxy- $\beta$ -D-glucopyranosyl) - (1 $\rightarrow$ 3) - *O* -  $\beta$  - D - galactopyranosyl - (1 $\rightarrow$ 4) -  $\beta$  - D - glucopyranosyl - (1 $\rightarrow$ 1) - (2*S*, 3*R*, 4*E*) - 2-tetracosanoylamino-4-octadecene-1,3-diol (2). (Sialyl dimeric Le<sup>a</sup> ganglioside). A solution of **41** (8 mg, 2.3  $\mu$ mol) in pyridine (1 mL) was added dropwise onto LiI (8.1 mg, 60.5  $\mu$ mol, dried at 200 °C for 1 day in vacuo), and the mixture was heated for 7 h at reflux under argon. The reaction mixture was chromatographed first on Sephadex LH-20 with 1 : 2 CHCl<sub>3</sub>-MeOH and then on SiO<sub>2</sub> with 10 : 1 CHCl<sub>3</sub>-MeOH to afford **42** (7 mg, 88%). A solution of **42** (7 mg, 2.0  $\mu$ mol) in EtOH (5 mL) was added MeNHNH<sub>2</sub> (2.5 mL), and the mixture was stirred for 18 h at 80 °C, and then concentrated in vacuo. The residue was purified over Sephadex LH-20 with 5 : 5 : 1 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O to afford the amino derivative. To the solution of amino derivative in 2 : 2 : 1 MeOH-THF-CH<sub>2</sub>Cl<sub>2</sub> (2.5 mL) was added acetic anhydride (100  $\mu$ L), and the mixture was stirred for 3 h at room temperature and concentrated in vacuo. The residue in 1 : 1 MeOH-THF (0.6 mL) was stirred with 1N NaOH (0.3 mL) for 1 day at room temperature, and the mixture was chromatographed on a Sephadex LH-20 with 5 : 5 : 1 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O to give **2** (3.4 mg, 74%): <sup>1</sup>H NMR (50 : 1 (CD<sub>3</sub>)<sub>2</sub>SO-D<sub>2</sub>O)  $\delta$  5.542 (dt, 1H,  $J = 7.0, 15.0$  Hz, H-5Cer), 5.354 (dd, 1H,  $J = 7.0, 15.4$  Hz, H-4Cer), 4.792 (br-d, 2H, H-1h and H-i), 4.782 (d, 1H,  $J = 8.1$  Hz, H-1c), 4.764 (d, 1H,  $J = 8.0$  Hz, H-1e), 4.621, 4.555 (2q, 2H, H-5h and H-5i), 4.362 (d, 2H,  $J = 8.0$  Hz, H-1d and H-1f), 4.270 (br-d, 1H, H-1b), 4.167 (d, 1H,  $J = 8.1$  Hz, H-1a), 2.754 (dd, 1H,  $J = 4.9, 12.1$  Hz, H-3 $eeq$ ), 1.879, 1.831, 1.819 (3s, 9H, 3NAc), 0.840 (t, 6H,  $J = 7.0$  Hz, 2CH<sub>2</sub>Me); FABMS (TEA matrix):  $m/z$  (M-Na)<sup>+</sup> 2287.**

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## REFERENCES AND NOTES

1. Dedicated to the memory of Professor Akira Hasegawa.
2. H. Koprowski, Z. Stepiewski, K. Mitchell, M. Herlyn, D. Herly and P. Fuhrer, *Somat. Cell Genet.*, **5**, 957 (1979).
3. S. Itali, J. Nishikata, T. Yoneda, K. Ohmori, S. Tsunekawa, N. Hiraiwa, H. Yamabe, S. Arii, T. Tobe and R. Kannagi, *Cancer*, **67**, 1576 (1991).
4. G. C. Hansson, *Trends in Glycoscience and Glycotechnology*, **9**, 46, 211 (1997).
5. A. Endo, M. Iida, S. Fujita, M. Numata, M. Sugimoto and S. Nunomura, *Carbohydr. Res.*, **270**, C9 (1995).
6. S. Nunomura, M. Iida, M. Numata, M. Sugimoto and T. Ogawa, *Carbohydr. Res.*, **263**, C1 (1994).
7. M. Iida, A. Endo, S. Fujita, M. Numata, K. Suzuki, S. Nunomura and T. Ogawa, *Glycoconjugate J.*, **13**, 203 (1996).
8. A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, *J. Carbohydr. Chem.*, **13**, 641 (1994).
9. M. Mori, Y. Ito and T. Ogawa, *Carbohydr. Res.*, **195**, 199 (1990).
10. a) K. Koike, Y. Nakahara and T. Ogawa, *Glycoconjugate J.*, **1**, 107 (1984);  
b) K. Koike, M. Numata, M. Sugimoto and T. Ogawa, *Carbohydr. Res.*, **158**, 113 (1986).
11. S. Sato, S. Nunomura, T. Nakano, Y. Ito and T. Ogawa, *Tetrahedron Lett.*, **29**, 4097 (1988).
12. a) S. Nunomura and T. Ogawa, *Tetrahedron Lett.*, **29**, 5681 (1988); b) S. Sato, S. Nunomura, T. Nakano, Y. Ito and T. Ogawa, *Tetrahedron Lett.*, **29**, 4097 (1988);  
c) S. Sato, Y. Ito and T. Ogawa, *Tetrahedron Lett.*, **29**, 5267 (1988).
13. a) L. M. Haines and E. Singleton, *J. Chem. Soc., Dalton. Trans.*, 1891 (1972);  
b) J. J. Oltvoort, C. A. A. Van Boeckel, J. H. De Koning and J. H. Van Boom, *Synthesis*, 305 (1981).
14. a) Wm. Rosenbrook Jr, D. A. Riley and P. A. Lartey, *Tetrahedron Lett.*, **26**, 3 (1985); b) G. H. Posner and S. R. Haines, *Tetrahedron Lett.*, **26**, 935 (1985).
15. a) O. Kanie, M. Kiso and A. Hasegawa, *J. Carbohydr. Chem.*, **7**, 501 (1988);  
b) A. Hasegawa, H. Ohki, T. Nagahama, H. Ishida and M. Kiso, *Carbohydr. Res.*, **212**, 277 (1991).
16. Y. Ito and T. Ogawa, *Tetrahedron Lett.*, **29**, 106 (1988).

17. O. Kanie, Y. Ito and T. Ogawa, *J. Am. Chem. Soc.*, **116**, 12073 (1994).
18. a) H. Paulsen and H. Tietz, *Angew. Chem. Int. Ed. Engl.*, **21**, 927 (1982); b) H. Paulsen and H. Tietz, *Carbohydr. Res.*, **125**, 47 (1984); c) K. Okamoto, T. Kondo and T. Goto, *Tetrahedron Lett.*, **27**, 5229 (1986).
19. T. Matsumoto, H. Maeta, K. Suzuki and G. Tsuchihashi, *Tetrahedron Lett.*, **29**, 3567 (1988).
20. G. Excoffier, D. Gagnaire and J-P. Utille, *Carbohydr. Res.*, **39**, 368 (1975).
21. R. R. Schmidt and J. Michel, *Angew. Chem. Int. Ed. Engl.*, **19**, 731 (1980).
22. a) H. Kunz and W. Sager, *Helv. Chem. Acta.*, **68**, 283 (1985); b) K. C. Nicolaou, A. Chucholowski, R. E. Dolle and J. L. Randall, *J. Chem. Soc., Chem. Commun.*, 1155 (1984).
23. W. S. Mungall, G. L. Greene, G. A. Heavner and R. L. Letsinger, *J. Org. Chem.*, **40**, 1659 (1975).
24. E. Bald, K. Saigo and T. Mukaiyama, *Chem. Lett.*, 1163 (1975).
25. a) E. Taschner and B. Liberek, *Rocz. Chem.*, **30**, 323 (1956); b) E. Taschner and B. Liberek, *Chem. Abst.*, **51**, 1039d (1957); c) F. Elsinger, J. Schreiberand and A. Eschenmoser, *Helv. Chim. Acta*, **43**, 113 (1966); d) M. Sugimoto, M. Numata, K. Koike, Y. Nakahara and T. Ogawa, *Carbohydr. Res.*, **156**, C1 (1988).
26. M. S. Motawia, J. Wengel, A. E. S. Abdel-Megid and E. B. Pedersen, *Synthesis*, 384 (1989).
27. R. Kuhn, P. Lutz and D. L. MacDonald, *Chem. Ber.*, **99**, 611 (1966).
28. a) B. Helferich, *Adv. Carbohydr. Chem. Biochem.*, **7**, 209 (1952); b) B. Helferich and J. Zirner, *Chem. Ber.*, **95**, 2604 (1962).
29. a) T. J. Martin and R. R. Schmidt, *Tetrahedron Lett.*, **41**, 6123 (1992); b) H. Paulsen, H. Tietz, *Carbohydr. Res.*, **125**, 47 (1984).
30. T. Mukaiyama, Y. Murai and S. Shoda, *Chem. Lett.*, 431 (1981).